



REVIEW ARTICLE

Microbial enzymes: the role of enzyme in cancer therapy

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Abstract

One of the most important challenges of the 21st decade is cancer. Adequate therapy advances aren't meeting the growing quantity of sufferers. Therapies that are frequently utilized doesn't always achieve the expected outcomes. As a consequence, it's vital to take action. Identify innovative supplementary appropriate therapies. Immunology involves the utilization of certain kinds of microorganisms. It is perhaps of their most essential potentially fruitful pathway that, in some way, therapy is thought to increase intestinal response, allowing cancerous lymphocytes to be removed preferentially. These study results seem encouraging, proving that microbial activation creates a strong reaction engagement in the innate immunological responses. Also, microbes may be used for various methods depending on their special qualities, such as pathogenicity, anaerobic living, and binding compounds that may be transported to a particular region. This publication gives an analysis of a chosen listing of indigenous microorganisms, including their characterization, genetic encoding, enzymes associated with proteins, mechanisms, or action towards cancer. Things are still in the testing stage. The use of microorganisms for chemotherapy, unlike every medical treatment, has inherent drawbacks. Biotechnology makes use of a large variety of enzymes that are commercially manufactured using specially screening microbes. Several microbes have been studied, developed, or optimized to create large enzymatic preparations for commercial production. Because diverse businesses need enzymes for diverse objectives, microbiological enzymes have been researched for their unique properties that may be used in a variety of bioreactors.

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1. Introduction

Annually, millions or even billions of people are affected by cancer. However, more than half of these people perish from complications of their condition worldwide. A total of 10.9 million primary cancer diagnoses (including melanoma) is recorded, with 6.7 million fatalities related to the illness T(GLOBOCAN). Cancer is accountable for roughly 1/5 of all deaths in the industrialized nations of the West. Conversely, one part of each 3 individuals will be confronted with such deadly cancer at some point throughout their life. Throughout

Europe, cancer has become a severe general populace wellness major worry, with a probability of around 3%, going to rise to 15% in older years. The growing impact is mostly due to an ageing populace, so it requires a concerted effort by oncologists, public health professionals, legislators, including scientists [1]. Surgical, chemotherapeutic, or radiation are quite frequent cancer therapies that are unable to reach full illness eradication. Additionally, it is well accepted that irradiation and electromagnetic fields may cause cancer. Chemotherapeutic will almost certainly result in serious negative impacts. As a result, several innovative methods for

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cancer therapy have been developed. However, the topical use of living, wounded microorganisms or their refined metabolites is just an instance.

Two different methods for therapy include local/regional therapy or evaluating the effectiveness or tolerability. Whenever various techniques are utilized simultaneously, it is called combo treatment. Regional therapy possibilities include brain metastases excision or radiotherapy. Surgical chemotherapy is critical in that treating cancer because it often gives the greatest chance towards achieving complete cure (radical treatment). It's also used in hospice therapy, which really doesn't give a treatment rather assists terminally sick people manage chronic illnesses or preserve optimum functioning during their next stages. Surgery significantly reduces the tumor size, which improves the efficiency of systemic treatment [2]. Some other kind for treatment is brachytherapy, which involves incinerating the tumor and causing a reduction in biological divisions and physiological processes. Radioactive treatment involves something like an exterior agency and the insertion of a generator inside and around a tumor. Standard treatment has a different impact dependent upon if the person's whole body is impacted. Chemo, hormonal treatment, or therapeutic strategies are essentially distinct treatments. Chemotherapeutic is a type of treatment in which drugs are used to prevent organisms from proliferating. Most quickly dividing cells, including cancerous or non-cancerous, are destroyed. Chemotherapeutic is linked to a wide range of hazardous reactions, and a steady decline in the patient's condition. Like a consequence, scientists were on the seek for potential, better targeted treatments. Hormonal therapy is used to cure malignancies which have oestrogen receptors, resembling those of the breast, ovary, and prostate. Despite the fact that this technique relies based on hormonal imbalances, it is crucial to check transcriptional activity before starting the procedure since they might vary as the tumor advances [3]. Recurring tumors are frequently treated with hormonal therapy. Recombinant human chemo is one greatest prevalent kind of pharmacological treatment. Allergens discovered in cancerous cells are being targeted by investigators. Monoclonal antibodies are often used to impair the metabolic pathways of cancerous cells. Chemotherapeutic treatment mainly includes immunization using well oligodendrocytes or malignant cells. Interestingly, microbial vaccinations have been employed like a tumor treatment for years to urge the person's antibodies to fight the disease; unfortunately, it sort of treatment still seems to be underexplored.

2. Type of Microbial Enzyme

2.1 Cytosine deaminase

Throughout fungi and bacteria, cytosine deaminase catalyzing the hydrolyzing demethylation of the purine basic cytosine (cytidine without the oligonucleotide element) producing

pyrimidines, but not in humans. Initially transformed to uracil and subsequently to the uridine monophosphate, this molecule acts like a nucleic acid biosynthesis substrate. In cytosine, *Escherichia coli* (*E. coli*) cytosine deaminase has $K_m = 0.22 \text{ mM}$ and $k_{cat} = 185 \text{ s}^{-1}$. During catalysis, the enzyme needs any metallic ion with two valences, so iron (II) may be eliminated since that too, producing an apoenzyme with only 5% of the original catalytic properties^[4]. With both the introduction with presence of chelating, metal, iron and zinc, copper, or the apoenzyme may be recreated. Although the K_m values for the specific enzymes are comparable, restoration using iron generates one of most active form, whereas zinc adding yields a $k_{cat} = 32 \text{ s}^{-1}$ enzyme. A functioning deaminase is inhibited by copper or zinc ions, as well as the enzymes have two metal binding affinities. Cu inhibits catalysis without displacing the catalytically important metal. Physical designs for microbial, fungal, or human nucleotides solution shows in this figure 1 that how two separate proteins fold evolved in parallel to perform the very same process on a wide range of substrate. As a result, the cytosine deaminase efficiency among both the bacteria and individuals indicated has been tested. Despite differences, all 5FC-resistant specimens, as well as 5FC-susceptible comparator strains 6936 or 5094, demonstrated significant cytosine e3 ligase function (Table 1).

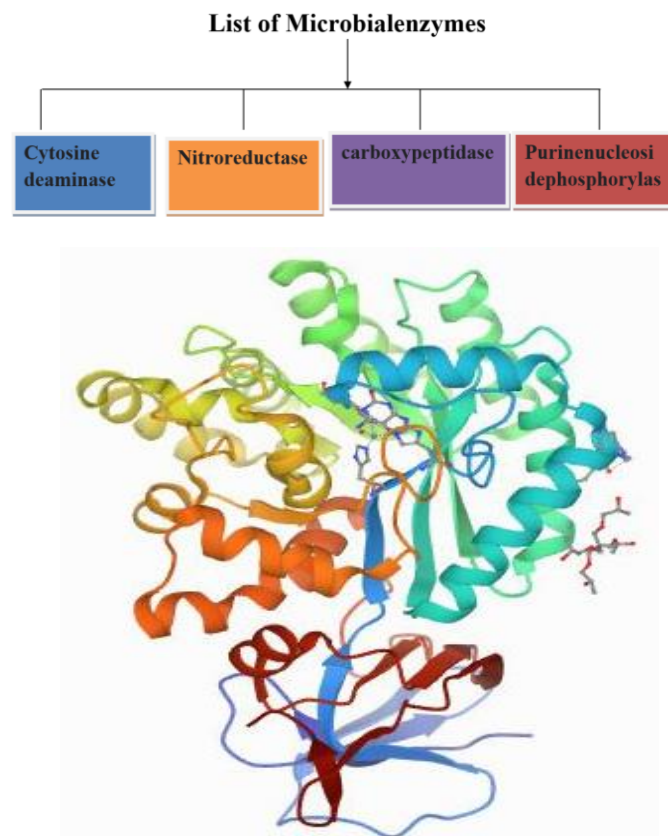


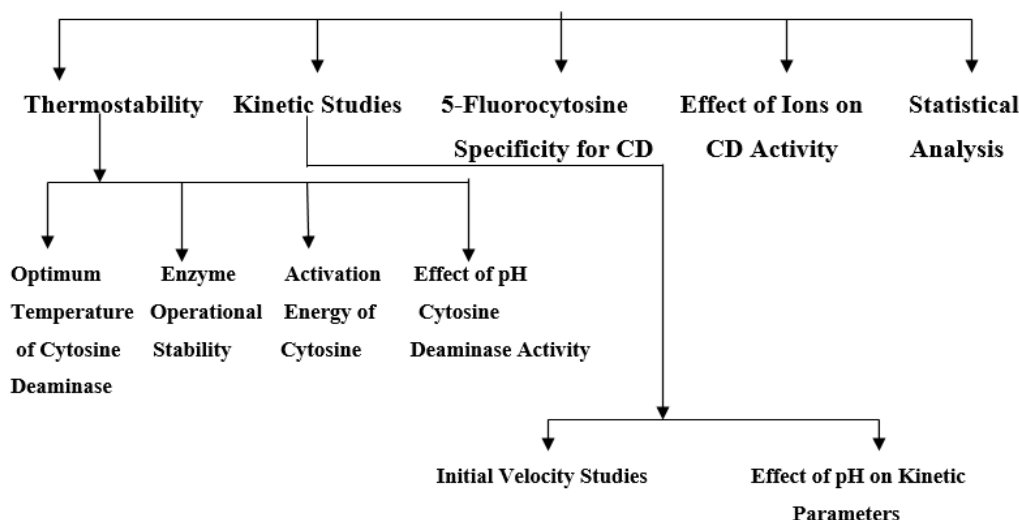
Figure 1: Crystal structure of Cytosine Deaminase

Table 1: Expressed as Nanomoles of deaminated cytosine per minute per milligram of protein; values are average of three independent assays.

Strain and isolate	MIC (mg/ml) of 5FU	Cytosine deaminase activity	K _t (app) Mm	V _{max} (nmol min ⁻¹ 10 ⁷ cells ⁻¹)
6936	≤0.5	7.89 ± 0.81	6.15 ± 0.18	1.20 ± 0.50
5094	≤0.5	20.98 ± 3.52	7.06 ± 3.03	1.09 ± 0.58
CL60	≥256	48.3 ± 2.79	10.78 ± 0.32	1.63 ± 0.64
CL31	≤0.5	17.38 ± 2.41	NM	NM
CL38	≤0.5	12.83 ± 4.34	NM	NM
CL42	≤0.5	25.15 ± 0.64	NM	NM
CLE11-801	≤0.5	7.40 ± 0.11	NM	NM

Regarding *C. lusitanae* 5FC-resistant isolates including variety, 5FU MICs, cytosine deaminase activity, or kinetics characteristics of [14C] 5FC absorption were determined.

2.2 Characterization of Cytosine Deaminase



2.3 Gene Coding of Cytosine Deaminase

Ligase culturing refers to the process of giving specific proteins to somatic mutations that are inadequate or weak in particular essential enzymes (for instance, CADases, adenosine kinase, and TK). Both procedures used one toxin, one detoxification enzyme, and one ligase protease complex [5]. These tactics don't function using CDase as no harmful molecule has been identified that CDase can preferentially detoxify.

In conclusion, when combined with PALA, cytosine, or inosine in the strong selective medium, the bacteria CDase genes may be employed as just an effective selection indicator. Positive selection is a secure and convenient way to manipulate cells that have been changed or have the bacterium CDase genes. When combined with the negative classification utilizing 5-FC, the positive selection enables employing the microbial CDase into patient anti-cancer therapies extremely successful or appealing.

2.4 Structure of Cytosine Deaminase protein

Literary hanging drop vapour diffusion has been used to increase *Escherichia coli* cytosine deaminase granules

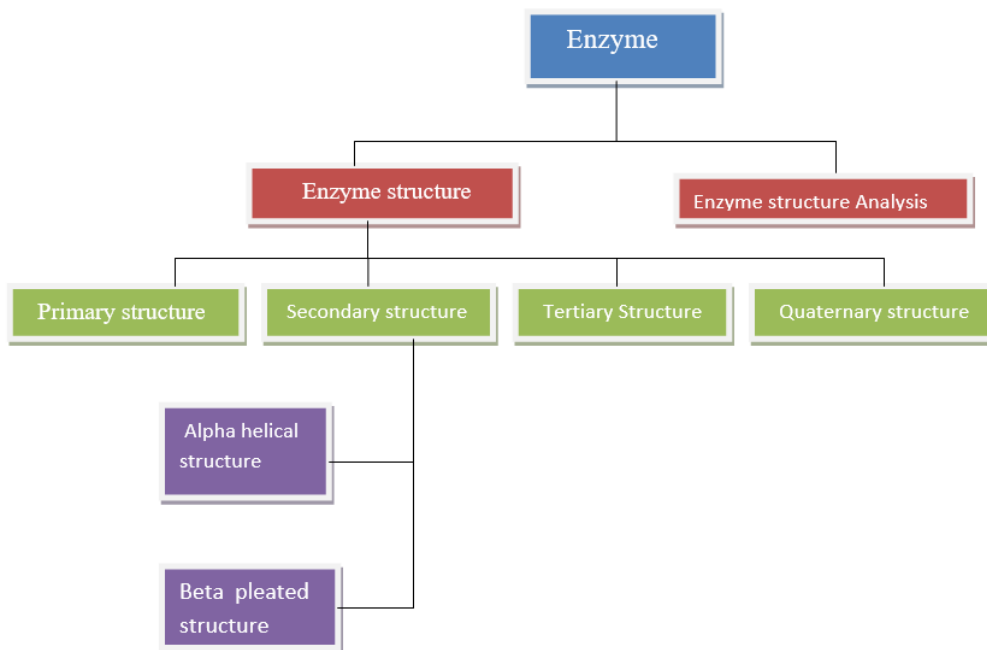
complexed with Zn²⁺ and phosphonocytosine by blending different quantities of nutrients and adsorbate and manually more than 1.0 mL of precipitating agent at ambient fever. This emulsion of proteins contains untamed CDA extracted from cells cultured inside 50 mM Tri's buffer, pH 7.5, 1.0 mM ZnCl₂, or 10 mM phosphonocytosine inside the addition of 1.0 mM Zinc chloride (18 mg/mL). 35 % pentaerythritol propoxylate, 0.05 M HEPES, pH 7.5, 0.2 M potassium chloride, plus 1.0 mM ZnCl₂ were present in the precipitating agent [6]. These crystals formed in eight and nine days to display diffraction gratings compatible with the R32 spatial grouping, including 1 peptide particle for each asymmetric subunit. Those crystals were immersed in cry-protectant mixtures prepared from native mom fluid and supplemented with 20% glycerol for data collection. Such crystals were illuminated inside the nitrogen stream after a 10-second incubation time. Refraction data was gathered on an ADSC CCD detector at the NSLS beamline (Brookhaven National Laboratory). The programs DENZO or SCALEPACK have been used to combine or time scale scattering intensity.

- Enzymes are proteins formed up of amino acids linked together within one or even multiple peptide chains. This

basic structure alludes with relation to the amino acid sequencing of polypeptides [7]. As a consequence, the 3D model was created. proteins' structural analysis, this shape of the peptide interface, for example, is specified.

- The four stages of specific enzymes are primary, secondary, tertiary, and quaternary.
- This The amino acids sequencing for any protein comprises the fundamental structure.
- Secondary structural refers to how amino acids combine inside a chain throughout enzymes.
- Helical (also known as helices) or pleated sheets (also known as pleated sheets) are the two forms of structural components.

- This Tertiary structure is the three-dimensional organization of amino acids.
- The connection between polypeptide subunits is referred to as quaternary structure.
- Powerful analysis techniques/equipment may be used to study enzymatic structure, including Amino acid detectors, peptide mappings, Edman disintegration, spectrometry, cyclic dichroism spectrometry, fluorescence spectroscopy, X-ray crystallography, or atomic magnetically resonant are all examples of techniques used to study proteins.



This configuration of an enzyme may vary from a simple to either a three-dimensional amino acid chain foldable protein complexes. The amino acid composition in protein molecules determined every enzyme's three-dimensional structure. Furthermore, the activity of an enzyme is determined by its shape.

2.5 Mechanism

Throughout a mixture of NMR or crystallographic investigations, the structural and mechanism of two different nucleoside deaminases, adenosine deaminase and cytidine deaminase, have been thoroughly described. 5 out of 10 Despite this fact that they both use a catalytic zinc ion or have comparable processes, the two enzymes have distinct structural folding or catalytically active layouts. Figure no. 2, depicts a working model for the deamination of cytosine reaction mechanism [8]. The composition of CDA inside the vicinity of a phosphonate inhibitor that binds tightly. As well

as mutagenesis tests show that this enzyme needs a solitary chelating ion in an active center for the organization of the cytosine-targeting water molecule. Again, for stimulation of this hydration that is nucleophilic and following translocation of protons events, 3 major region residues are also needed. Glu-217, His-246, and Asp-313 are among these residues [9]. This catalytic effectiveness of CDA is drastically reduced when either of those sites is mutated. Adenosine deaminase, SAH deaminase, guanine deaminase, soxanthopterin deaminase, and oxoguanine deaminase, isoxanthopterin deaminase all have the same trio of characters in their active sites. Throughout the procedure explained above, every metal-bound molecule of water is chemically bonded to Asp-313 and His-246. Inside the lack of a substrate's counterpart, the X-ray structural of CDA (PDB code: 1K6W) supports it [10]. This becomes obvious from the X-ray spectra for While with the existence of CDA of inhibitors 2 (PDB code: 1K70) or 5 (PDB code: 3O7U) that Glu-217 is situated to supply protons towards the output uracil's N3 region. Unfortunately,

this remnant seems to be enough away from a water molecule linked to a metallic to allow straight protons extraction. This propensity of the E217A mutation to catalyze Compound 4 is hydrolyzed. Without oxygen is protonated. Placed in the corresponding predicament of N3 additional supports the function of Glu-217 suggested herein. Inside the hypothesized process, a proton is transported from of the metal-bound water molecule to Glu-217 via His-246. This is unclear if this

protonation happens before or after cytosine binds to the active center. Metals coupled hydroxide assaults the re-face and cytosine after the substrates is attached inside the protein surface. The molecular composition of inhibitory 2 in the bind catalytic place of Fe-CDA or inhibitors 5 linked to Zn-active CDA's site supports the stereochemistry of such an event.

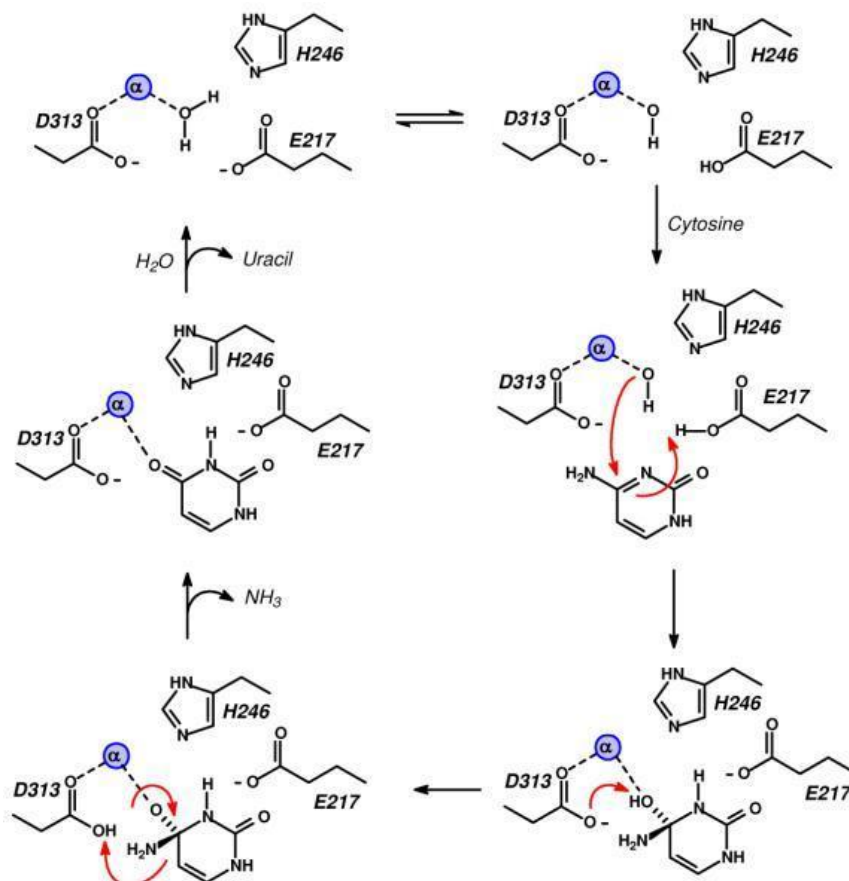


Figure 2: Mechanism for Cytosine deaminase

2.6 The Cytosine Deaminase Enzyme Against cancer

Recombinant prodrug 5FC has a dosages relationship. Higher 5FC concentrations resulted in more CD-expressing cells being killed fractionally. That dosage impact of CD gene expression is less obvious, as the cellular line's innate susceptibility may eventually determine it to 5FU. This amount of increased CD expression seems to be related to cytotoxicity in sensitive cell cultures like colorectal, breast, or lung cancer [11]. A CD dose-response relationship is unclear in less sensitive sarcoma cell cultures. Cytosine deaminase/5-fluorocytosine (CD/5-FC), which was investigated extensively over the last year, is among the most intriguing molecular chemotherapeutic approaches. CD is a bacterium or fungal protein capable of converting the antifungal agent 5-FC to the chemotherapeutic drug 5-fluorouracil (5-FU). CD is

found inside prokaryotes and fungi, though not in upper eukaryotes, wherein it is a key component of the pyrimidine salvaging process. 5-FC, an antimicrobial drug that needs the stimulation of the fungus CD, is widely employed to treat Cryptococcus or Candida infections in the prior [12]. As a result, in terms of toxicological characteristics people are well-known. 5-FU is a pyrimidine antagonist wherein fluoride replaces the hydrogen atom at the C-5 site of pyrimidines. Because 5-FU has a comparable framework to uracil, this can be transformed molecules that are physiologically active like 5-fluoro-2'-deoxyuridine-5'-monophosphate (FdUMP) or 5-fluorouridine-5'-triphosphate, as well as metabolized to motionless precursors like 5-fluoro-alanine, ammonia, and CO₂ via the identical anabolic and catabolic paths as pyrimidines. FdUMP forms a covalent ternary complex with thymidylate synthase (TS), a crucial DNA de novo synthetic

protease that catalysis the methylation of deoxy uridine monophosphate (dUMP) to deoxythymidine monophosphate [13]. 5-Fluorouracil triphosphate has been integrated in to the RNA as well as significantly interfere with the RNA manufacturing process, whereas FdUMP with the coenzyme N5, N10-methylene (dTMP). That prevents dUMP from being converted to dTMP and, as direct consequence, nucleotide pools for dTMP, a thymidine precursor in DNA synthesis, from being disrupted.

Nearly 2 centuries ago, that codA gene expressing bacterium CD (bCD) were translated from *Escherichia coli* into eukaryotic expression systems for GDEPT and mouse fibroblasts throughout Vivo or in murine tumor types (Mullen, Coale, Lowe, & Blaese).

3. Nitroreductase Enzyme

Like a suicidal gene therapy technique, an *Escherichia coli* protein nitroreductase (NTR) is combined with both the drug

carrier CB1954 [5-(aziridin-1-yl)-2,4-dinitrobenzamide]. CB is a mild anticancer drug that has been produced (Khan and Ross.). DT-diaphorase (NAD (P) H dehydrogenase) seems to be the protein that activates CB in mammals. CB1954 is converted to its 4-hydroxylamino derivative by this enzyme [14]. In a subsequent activation step, an alkylating chemical was created following acetylation through thioesters like co - enzyme A (CoA). This active synthetic opiate may then produce DNA cross-links that are difficult to repair. Recombinant *Escherichia coli* NTR is sensitive to CB1954, but human's DT-diaphorase is incapable of converting it, reducing the lethality of converted cells. The NTR/CB1954 method offers the benefit that death mediated by reactivated CB1954 is not reliant on cellular stage of the cycle, effectively allowing for the mortality of dormant cancer cells. See (Figure 3) This rhodamine-based luminous sensor has been used to image nitroreductase enzymes in living cells and also hypoxic.

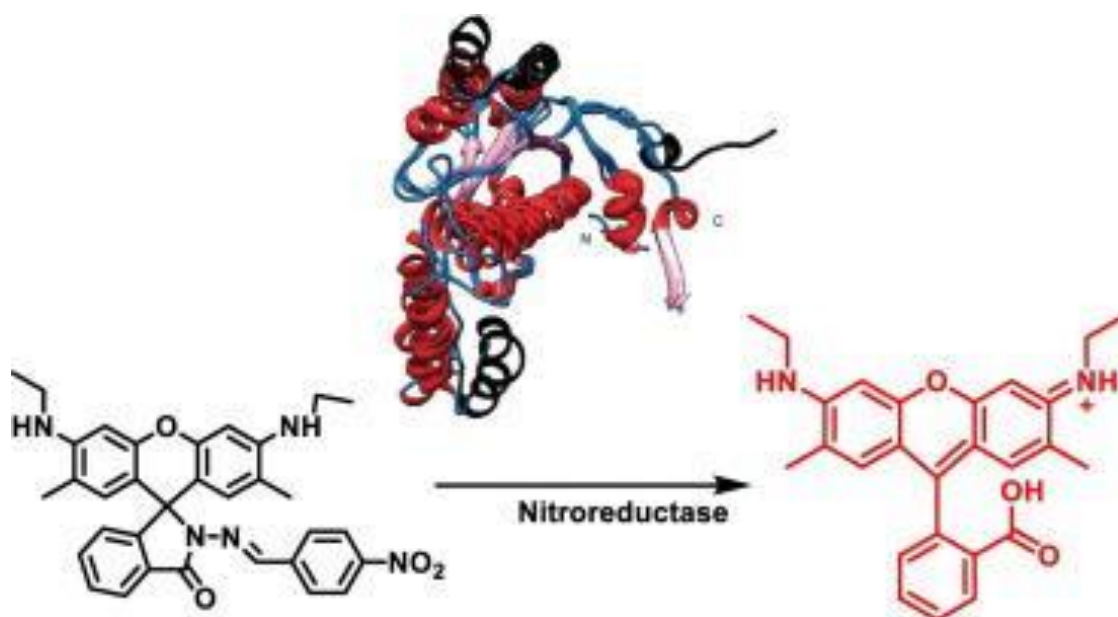


Figure 3: living Nitroreductase

3.1. Characterization

- We synthesised or analysed N-heterocyclic nitro derivatives (NHN1-16).
- Chromatography testing was used to determine these substances' Ssap-NtrBnitroreductase activity.
- Hep3B, PC3, or HUVEC cell cultures have been used to test the cytotoxic impacts of NHN1-16.
- The SRB test was used to evaluate 5 non-toxic chemicals with drug carrier potential.
- On PC3, the IC50 values for non-toxic chemical metabolites ranged from 1.71-4.72 nM.
- Drug carrier possibilities include the innovative pyrazole compounds NHN12 and NHN14.

3.2. Gene Coding of Nitroreductase Enzyme

GDEPT (gene-directed enzymatic drug carrier treatment) combines the use of cancer gene transfer vectors with systemic injection of ordinarily inert active drugs to produce extremely selective tumor-cell death. GDEPT might benefit from nitroaromatic active drug like CB1954, which are easily converted to strong DNA chemotherapeutic drugs by microbial nitroreductase enzymes (NTRs). This nfsB gene from *Escherichia coli* may boost tumor cell responsiveness to CB1954 by more than 1000-fold when transfected. Unfortunately, [15-16], the stimulation of CB1954 by NfsB at clinically relevant dosages is limited due to weak catalytic effectiveness. A shortage of versatile, elevated testing

methods has hampered attempts to find improved NTR alternatives. Using SOS chromo test and supplementary testing methods are utilized to assess new enzymes that react CB1954 as well as similar bio reductive and genotoxic derivatives in this paper. This primary *Escherichia coli* NTR, NfsA, is shown to be 10-fold more effective than NfsB in activation CB1954 the purified protein (kcat/Km) or as excessively in an *Escherichia coli* strain lacking the NfsA/nfsB genes. Consistently expressed in HCT-116 conventional colon cancer cells, NfsA provides susceptibility with CB1954 also with a comparable efficacy to NfsB. Researchers also discover 2 additional *Escherichia coli* NTRs, AzoR or Nema, which have never been studied before in the framework of nitroaromatic drug carrier activating.

3.3 Structure of Nitroreductase protein

The flavin nitroreductase from *Escherichia coli* eliminates a wide range with quinoline or nitroaromatic substrates. Its capacity to transform generally similar products that aren't harmful like CB1954 (5-aziridin-1-yl-2,4-dinitrobenzamide) into extremely cytotoxic compounds has sparked attention in its application to cancer genes treatment [17]. Researchers used 3 crystalline formations with resolutions of 1.7, 1.8, and 2.4, account for about 10 non-crystallographically connected monomers, to identify the structure of the protein linked to a substrate analog, nicotinic acid. This enzyme is dimeric, with a massive hydrophobic center, so each part comprises a five-stranded β -sheet encircled by α -helices. Every monomer's core region has helices E and F protruding from that too. The 15 C-terminal residues stretch all surrounding monomers, giving the 5th strands, so a large dimer contact is here. These adsorption sites are in solvent-exposed crevasses at the dimer interface, on opposite sides of the molecule [18-19]. The FMN establishes hydrogen bonds with one molecule as well as hydrophobic interactions with the other two, so its surface is hidden. Having only one hydrogen bonding between such protein, one nicotinic acid accumulates between both the dorsal surface of the FMN as Phe124 in helix F. This C4 atom of the coenzyme NAD(P)H would've been better placed for straight hydride transference onto flavin N5 only if nicotinamide circle remained at the same configuration as nicotinamide ligand. Compared to un-liganded flavin reductase and NTR, this complex contains helices E and F that are less mobile when ligands are bound. This structure clarifies the enzyme's broad substrate selectivity also offers the foundation for reasonable drug carrier development or location mutagenesis towards better enzymatic performance.

3.4 Mechanism

According to research, this same nitro group of CB1954 is converted to a hydroxylamine by 2 sequential two-electron exchanges. Even though the enzymes should accomplish the reducing of the nitrogen atom to a hazardous nitroso phase, NTR also isn't needed for the conversion of the nitroso

transitional to hydroxylamine. Like a result, our research work focuses just on enzyme-catalyzed conversion of CB1954's 4-nitro compound to a nitroso collective [20]. Inclusion of two protons plus two electrons is required to convert nitro to nitroso. The electrons should originate from the FMN component of NTR, and at least several protons should emerge from solutions, the second proton can originate through FMN or remedy. The coordinated transmission a single proton plus 2 charged particles with the same source, inside this instance the FMN, is typically referred known the hydride transmission. Pre or post the hydride exchange, a proton in solutions (for just a maximum of 2 protons + pair of electrons) could be added. This hydride may receive two electron acceptors: this same nitrogen firm's nitrogen and or one of the other oxygens. Although hydride transference to nitrogen necessitates further rearranging to generate the nitroso, the electro positive character of nitrogen makes it an effective receiver for such negatives hydride ions [21-22]. For hydride transference to occur, a single nitro firm's nitrogen or oxygen molecules should come into touch with the HN5 of FMN through van der Waals touch. As all these the HOMO (highest occupied orbital) of FMN as well as the LUMO (lowest unoccupied orbital) of CB1954 are dispersed over a lot of one's corresponding particles, transfer of an electron (whereby the charged particles emerge from FMN although both charged particles finally came from solution) also necessitates that only some part of CB1954 has been now within van der Waals interaction among some portion of FMN, whereas the octane organization is revealed to real fix. Experimental facts support either hydrogen transference or electrons exchange. Hydride transference has been thought may become implicated for the first substantial reduction to FMN via NAD(P)H.

3.5 The Role of Nitroreductase Enzyme Against cancer

Aromatic nitro compounds are converted to their respective amino equivalents by nitroreductases (NTR), however the equivalent catalysts in human organisms have yet to be identified and characterized. Mammalian NTR function, on either hand, has indeed been touted as a promising candidate again in the discovery of theranostic medicines for cancer or tumoral hypoxic [23]. Tiny molecule-based fluorescent dyes have recently emerged as an effective technique for detecting NTR activation. Furthermore, assessing NTR activities across various cells has been limited because most sensors rely on fluorescent fluctuations influenced by both enzymatic and non - enzymatic variables.

4. Carboxypeptidase (E) Enzyme

This enzyme was first extracted in bovine adrenal chromaffin granules like a carboxypeptidase B-like exopeptidase, wherein it has been Enkephalin precursors' C-terminal fundamental groups demonstrated to be removed [24-25]. The CPE is available in both liquid and transmembrane forms

and contains its pH of 5.5. This shape, with such has a molar weight of 53 kDa, is indeed a metabolic protein which, after peptidase dissociation, removes C-terminal lysine or arginine residue from neuropeptide precursors to create bioactive hormone as well as neuropeptides. The 55 kDa transmembrane form, on either hand, functions as a sorted receptor, directing a variety of prohormones or proneuropeptides to a controlled secretion route (RSP). Figure 4, depicts CPE from its structure to its functioning or illness, with an emphasis on the novel functions that even this unusual enzyme performs in a range of biological processes. CPE finally cleaves such mixtures to consider removing the fundamental contaminants. Whereas the activity of the proprotein automated machines with CPE is sufficient to produce this pharmacological peptide for several sequences, increased computational processes, like C-terminal acetylation, are necessary for the other proteins to form the reactive polypeptide.

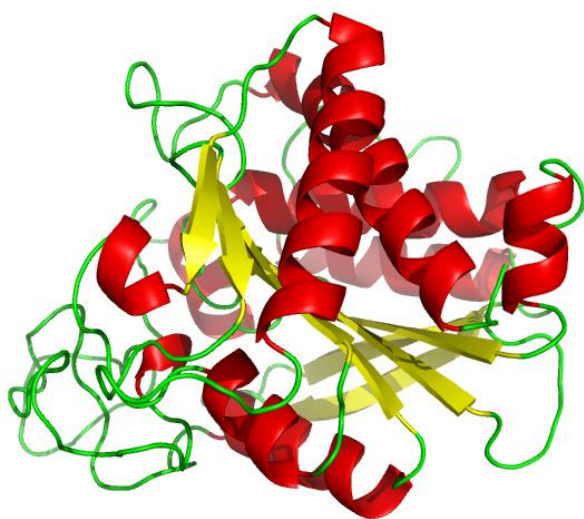


Figure 4: Carboxypeptidase (E) Enzyme

4.1 Characterization

Thus far, CPE was only biologically active defined within few animals. This was initially synthesized from bovine adrenal chromaffin granules then investigated extensively in mice (*Mus musculus*) or rats (*Rattus rattus*) (*Rattus norvegicus*). Manser was the earliest one to identify human CPE. CPE has also been investigated in chicken (*Gallus gallus*), thymus, or nematode tissues [26-27]. CPE was later discovered inside this genomic sequences of a chimpanzees (*Pan troglodytes*), monkey (*Macaca fascicularis*), swine (*Sus scrofa*), clawed frog (*Xenopus tropicalis*), sea-slug (*Aplysia californica*), or zebrafish via blast analysis as well as data gathering (*Danio rerio*). CPE has a surprising level of protein homology across phyla, indicating that it had a physiological consequence early in evolutionary.

4.2 Gene Coding of Carboxypeptidase (E) Enzyme

The nucleotide sequencing cytogenetic enzyme of *Pseudomonas* that encodes the carboxypeptidase G2 (CPG2) protein has been identified. Through matching the anticipated along containing amino acid of arbitrarily produced peptide fragments and so by N-terminal decoding the protein's sample, the genetic code was validated. Being in N-terminus, this gene codes for a 22-amino-acid signal sequence that closely mimics the signaling peptides from the other associated proteins [28]. A 36-amino-acid signal peptide has also been discovered that also might act with *Pseudomonas*. This gene's translational use is impacted by the DNA's high G + C (67.2 %) composition, with a 92.8 % predilection for codons culminating in G or C. This unique codon preference might play a role inside the normally expression level of *Pseudomonas* genes in *Escherichia coli*. The ribosome binding site or two potential promoters' sequences was discovered inside a stretch of DNA downstream of the gene encoding.

4.3 Structure of Carboxypeptidase E Protein

A genetic material was shown to include genetic traits for Zn-carboxypeptidase triad enzymes. Just as cellular levels, CPE has 21% similarity mostly with carboxypeptidase A (A1, A2, A3, A4, A5, and A6) and carboxypeptidase B (B1 and B2), including 55 and 37 % citrix, correspondingly [29-30]. Carboxypeptidase O has just 32% sequence coverage and is 22 % similar now at cellular stages. Furthermore, carboxypeptidase M or carboxypeptidase N1 were 46 % similar just at enzyme expression having 82 % structural saturation and 51 % similar just at amino stage having 86 % sequencing content, respectively. Carboxypeptidase D, for example, contains numerous Zn-carboxypeptidase domains and is 50% similar just at proteins layer, with 86 % sequencing identity. Beyond the Zn-carboxypeptidase domain, CPE and CPA/B proteins have relatively little to nothing in common, indicating that such molecules separated incredibly early in development [31]. Several taxa contain haplotypes sequence data of the Cpe gene, spanning the evolutionary tree from invertebrates Protostomia (Ecdysozoa, nematodes) through vertebrates Deuterostomia. An overall reducing graph depicts basic evolutionary connections based upon protein pattern similarity.

4.4 Mechanism

- According to more on their activity area mechanism, carboxypeptidases were typically divided within each of many groups.
- Carboxypeptidases were enzymes that have a metallic with inactivity cent (EC number 3.4.17).

- Serine carboxypeptidases include carboxypeptidases that contain serine groups within the working centre (EC number 3.4.16).
- Cysteine Carboxypeptidase E (or Carboxypeptidase) enzymes utilize enzymes as the protein surface (EC number 3.4.18).
- Those designations have nothing to do with the selection of the liberated glutamic acids.

4.5 The Role of Carboxypeptidase(E) against Cancer

CPE has been demonstrated to improve cell viability in pheochromocytoma (PC12) and hepato cellular carcinoma (MHCC97H) cells under nutritional restriction and hypoxic conditions, but not cell growth. Through the ERK signaling pathway, CPE increases the synthesis of pro-survival/anti-apoptotic proteins such BCL-2 in MHCC97H cells. CPE promotes cell proliferation by promoting ERK up regulation [32]. CPE also inhibits wnt transmission. CPE limits cellular multiplication by interrupting Wnt signalling or decreasing the production of β -catenin, which plays an essential role in cancer cell proliferation, meaning that CPE prevents Wnt signalling. Amplification of CPE stimulates cellular growth in gliomas, showing it has a top player influence in tumors, according to further studies. CPE expression is similarly suppressed by hypoxia and carbohydrate restriction.

As a consequence, in the face of caloric restriction and hypoxia, the quantity of CPE available to drive PC12 and MHCC97H cell development may be minimal but adequate to safeguard a person from metabolism. This basic technique through which CPE regulates cell proliferation has to be moreover researched [33]. CPE may be able to regulate an additional method to get around it effects of β -catenin, resulting in a net effect of CPE that stimulates Proliferation of cells CPE has been discovered to decrease fibrosarcoma (HT1080) penetration and metastasis, as well as delay and impair glioblastoma cellular metabolism.

Studies on brain metastases, small lung malignancies, and large neuropsychiatric squamous cell carcinoma provide further evidence that high levels of CPE transcription indicate a good prognosis. CPE method of anti-migration and anti-proliferation remains unclear. Several of these suggested routes are tied to the Wnt signaling system, which regulates cell migration via several of its components' β -CPE was located in its center of hepatocellular carcinoma (HCC) cell cultures, in which it binds to chromatin demethylase 1/2 to activate the production of neuronal progenitor NEDD9 (progressively dysregulated proteins)-expressing cells [34-35]. Because NEDD9 enhances cell division to growth migration it's conceivable because in vitro the N-CPE may help tumors grow and migrate by producing the protein. By increasing β -catenin levels, N-CPE, as a favorable regulator of the Signaling pathway, might possibly increase cell proliferation.

Another method to consider for N-CPE-induced

tumorexpansion of metastasis is the commencement of NF- κ B transcription. This protein affects a variety of cancers and plays a role in their origin and recurrence. CPE amplification was seen elevated NF- κ B expression in pancreatic cancer, whereas CPE knockdown reduced NF- κ B production and hampered growth of cells and migration, indicating whether CPE controls pancreatic cancer by NF- κ B. Though all scale of N-CPE/CPE was not among them. Distinguished throughout the experiment; N-CPE, instead of comprehensive CPE, is expected to have a role in pancreatic cancer development or penetration. Furthermore, N-CPE has been connected to lung cancer tumor recurrence. Further study is required to validate N-involvement. Expression's CPEs in cancer development, infiltration, metastasis, and survival in numerous malignancies.

5. Purine Nucleoside Phosphorylase (PNP)

PNP is just a hexameric protein of *Escherichia coli* that catalysis the irreversible 6-amino and 6-oxopurine (2'-deoxy) ribonucleosides are phosphorylyzed to produce a critical boost or (2'-deoxy) ribose-1-phosphate. Avian or mammalian PNPs, on the other hand, are trimeric but only take 6-oxopurine nucleosides as a source [36]. The fact that such different catalysts have different particularities was used in genetic therapeutic approaches where some derivatives are degraded by *Escherichia coli* PNP never by the humans' enzymes. Despite catalyzing the very identical underlying biochemical process, individual amino acid compositions of both the tetramers but also hexameric PNPs were completely different [37]. A physical study of the catalyst surface of human and *Escherichia coli* PNPs may give a better groundwork for the growth of possible *Escherichia coli* PNP-specific drug candidates. Figure no. 5, Purine nucleoside phosphorylase is shown as a ribbon.

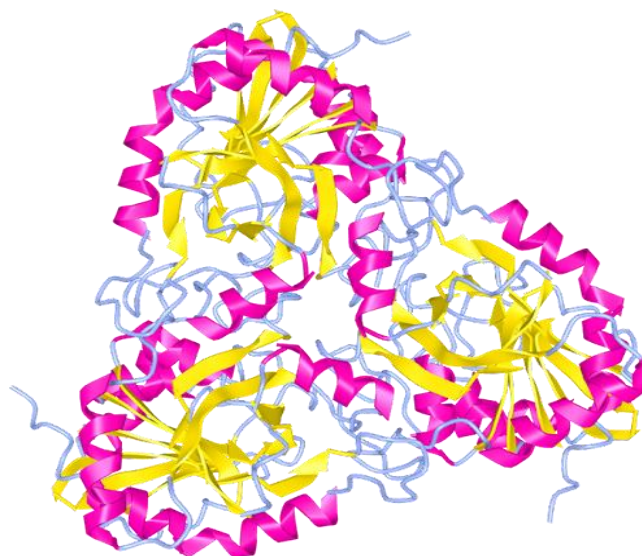


Figure 5: Structure of purine nucleoside phosphorylase

5.1 Characterization

P. falciparum were effectively cultivated to an 8 % aparasitemic rate using purine nucleoside phosphorylase-deficient human erythrocytes and study its development of properties in humans' infections purine nucleoside phosphorylase. Purine nucleoside phosphorylase production remained detected in untreated enzyme-deficient human red cells, however, increased to 1.5 % of the original erythron levels following parasitic infestation [38-39]. The parasitic purine nucleoside phosphorylase had an estimated natural molecular mass substrate specificity for inosine, guanosine, and deoxyguanosine but just not xanthopsin or adenosine, as well as a single major electrophoretic phase of pI 5.4.

The Km estimates again for fuels inosine or guanosine is four times smaller than the human erythrocyte enzyme. Researchers discovered two new powerful compounds in such trials 8-amino-5 deoxy-5 chloroguanosine and 8-amino-9-benzylguanine are purine nucleoside phosphorylase antagonists in both human erythrocytes and parasites [40-51]. Certain enzyme antagonists could offer modest antibacterial effect by decreasing hypoxanthine formation in parasitic organism erythrocytes.

5.2 Gene Coding of Purine nucleoside phosphorylase (PNP)

That genome produces an enzymatic that catalysis the phosphorolysis of purine nucleosides in a reversible manner [52]. The protein is trimeric, with triple components that are all similar. Impaired T-cell (cell-mediated) immunology is a consequence of nucleoside phosphorylase insufficiency, but it may also impact B-cell defense and antibody reactions [53]. Immune-compromised people may have neurological problems as well. This had been documented a recognized polymorphism in a position 51 which seems to have no effect on enzyme activity. On chromosomes, a pseudogene has been discovered [54-56]. One such gene codes for an enzyme that phosphorolyzes purine nucleosides in a reversible manner. Having three fragments make up the tetrameric enzyme. T-cell (cell-mediated) immunisation is harmed by mutations that cause nucleoside phosphorylase deficiency, as well as though B-cell immunity or immunoglobulin reactions could also be affected [57]. Throughout patient populations with immune deficiencies, neurologic disorders may appear. A genetic variation at a position 51 has been characterized which has no effect on enzymatic activity. Here on chromosome, a pseudogene has been found.

5.3 Structure of Purine nucleoside phosphorylase (PNP)

PNP Are utilizing congruous (56 % sequencing similarity) *Escherichia coli* PNP structural (PDB codes as just a suitable condition, this same system is found via molecule substitution in MOLREP (Vagin Teplyakov,). Across both rotational and translational computations, information with in variant 0–3 was considered, producing during an apparent solution having

a considerable difference, culminating in 6 compounds inside a symmetrical unit as well as a fluid concentration of 58.4 %. Their R factor was 45.7 as well as the relationship was 59.6%. Percent (R free = 45.6 %) after such a strict modification using REFMAC. REFMAC was used to further structural refining, and QUANTA (Accelrys Inc., San Diego, CA, USA) and COOT (Emsley's Cowtan) were used to develop models [58-59]. The ultimate R value for the system, which has It is 18.4 % (R free = 23.5 %) with 1399 protein acid repeats and 716, water molecules.

5.4 Mechanism

Purine nucleoside phosphorylase enzymatic performance was investigated using X-ray crystallography, molecular modelling, and location mutation (PNP). PNP catalyzing the phosphorolysis of purine nucleosides to pyrimidines and ribose 1-phosphate in a controllable way. with the use of a surface catalytic site [61-63]. An oxocarbenium ions is stabilized by laid phosphorus uses strong imagery, which works is major part of a catalytic triad in the suggested leaving group (TS) (Glu89-His86-PO4). Phosphate involvement in the TS is thought to be the molecular property which distinguishes phosphorylases other glycosidases as well as explains for PNP low hydrolytic performance [64-66]. A hydroxyl group among N7 and a conserved ASN is also included in the suggested PNP TS. The adverse charged that builds on the nucleotide ring after glycosidic bond breakage is stabilized by hydrogen bond transfer towards N7 within TS. The hydrogen bond was further supported by kinetic investigations employing N7-modified analogues [67-69]. PNP employs a ligand-induced conformational shift to place ASN as well as other critical proteins inside the active center for catalytic, according to crystallography analyses on 13humans Nelligan compounds. Purine nucleosides bind to PNP with a non - standard glycosyltorsional orientation (+anticlinal) and a rare sugar pucker (C4 endo), according to these findings.

The bonding configuration to improve phosphorolysis via ligands strain were expected by single goal calculation. Purine binding seems to come before ribose 1-phosphate binding inside the synthesis side, although the sequence of complex formation for key functions is more obvious [70-72]. The retention of catalyst has subsequently supported the hypothesized catalytic mechanism critical sites across nucleoside phosphorylases and selectivity towards 6-oxopurine nucleosides.

5.5 The role of Purine nucleoside phosphorylase (PNP) against Cancer

A purine rescue enzymes Purine nucleoside phosphorylase (PNP) transforms structure that defines purine and inositol to hypoxanthine. Methods: 279 samples from patients with various malignancies were obtained throughout therapy and matched to the regular community regarding serum PNP

activities all for pre/post phases [73-75]. Findings: In comparison to cancer individuals (pre-dose 12.37.4 U/l [n=215] and post-dose 11.25.9 U/l [n=64], plasma PNP activities were disclosed to just be 3.21.4 U/l (n=55). Serum PNP values did not vary significantly across cancer types but were on average four times higher than those seen in the overall community.

6. Conclusions

Microbial enzymes have enormous medicinal potential and are very important. This manufacture of therapeutic enzymes by bacteria is both cost-effective and environmentally benign. Those who may be useful in the medication of a range of human abnormalities, including the use of fibrinolytic enzymatic in the planning of myocardial injury, respiratory thrombosis, as well as venous cerebrovascular disease, enzymotics inside the regulation of infectious diseases, amino acid demeaning enzymatic through treating cancer, anti-inflammatory enzymes in scar as well as inflammatory diagnosis, as well as gastrointestinal aids inside the simpler gastrointestinal tract process. That requirement of the moment is to evaluate novel enzymes and to upgrade or enhance current ones. As enjoy the advantages of these enhanced microbial biopharmaceuticals over conventional chemotherapeutic drugs particularly in the therapy of many human illnesses, their isolation, control, oversupply, or uses of such enhanced microbial biopharmaceuticals must be investigated to their full capabilities. Humans have understood about enzymes since the dawn of civilization. Enzymes have been used extensively in a variety of industries, including ancient brewing as well as other applications. However, from the 18th century, it has been referred to as enzymes. Many scientists had attempted to investigate the utilization of enzymes, so we have learned about its prominence in our everyday lives as a result of their groundbreaking work. Nowadays, enzymes like: Cytosine Deaminase, Nitroreductase, Carboxypeptidase, or Purine Nucleoside Phosphorylase are chemically used for their critical life-saving medicinal applications. Ever since, intensive testing has led to the discovery of several microbes including its proteins having specific functions, which are today widely employed inside a range of manufacturing and medicinal domains. Their research and production of these therapeutically important enzymes has been at minimum as thorough as that of biocatalysts, highlighting the industry's potential near-term advantages. Chemotherapeutic therapy relying on microorganisms is usually neglected or underestimated. These tiny group of academics are aiming to find or improve microbe-based tumor treatment options, whether as vaccines which activate the immunological function to battle sickness or as carriers enabling the spread of anticancer medications.

Microbial sources are intriguing solid acid catalysts that have gotten a lot of interest because of their advantages above heterogeneous catalysis, like higher sensitivity and the

flexibility to work in mild processing circumstances. Additionally, several sectors need catalysts with specific features for application in the processing of therapeutic targeting or environmental assets. -glucosidases generated from *Fusarium* strains serve a crucial role throughout this respect owing to its specificity for just a wide spectrum of glycosyl substrates.

Despite the many production or purification issues which remain still been fully resolved in order to generate similar proteins on a commercial scale, -glucosidase has already been allocated to something like a number of key commercial processes owing towards its potential or broad activity. Basic importance of glucosidases and its production from *Aspergillus* strains is described in this article, as well as the main procedure variables, ingredients used, the quantities produced by submerged or vigorous fermentation. Purification, characterization, the spinal immobilization of -glucosidases, which are essential but potential catalyst inside a broad variety of industries, such gastronomic, pharmaceutical, medicinal, or renewables, were also analyzed throughout this section.

References

- [1] Ahmadpour, S., Hosseinimehr, S.J., 2018. PASylation as a Powerful Technology for Improving the Pharmacokinetic Properties of Biopharmaceuticals. *Curr. Drug Deliv.* 15, 331–341.
- [2] Alipour, H., Raz, A., Zakeri, S., Djadid, N.D., 2016. Therapeutic applications of collagenase (metalloproteases): A review. *Asian Pac. J. Trop. Biomed.* 6, 975–981.
- [3] Alkaade, S., Vareedayah, A.A., 2017. A primer on exocrine pancreatic insufficiency, fat malabsorption, and fatty acid abnormalities. *Am. J. Managed Care* 23, S203–S209.
- [4] Aneja, R., Datt, M., Yadav, S., Sahni, G., 2013. Multiple exosites distributed across the three domains of streptokinase co-operate to generate high catalytic rates in the streptokinase–plasmin activator complex. *Biochemistry* 52, 8957–8968.
- [5] Banerjee, A., Chisti, Y., Banerjee, U.C., 2004. Streptokinase—a clinically useful thrombolytic agent. *Biotechnol. Adv.* 22, 287–307.
- [6] Bassetto, F., Maschio, N., Abatangelo, G., Zavan, B., Scarpa, C., Vindigni, V., 2016. Collagenase From *Vibrio alginolyticus* Cultures: Experimental Study and Clinical Perspectives. *Surgical Innov.* 23, 557–562.
- [7] Basso, A., Serban, S., 2020. Overview of Immobilized Enzymes' Applications in Pharmaceutical, Chemical, and Food Industry. *Methods Mol. Biol.* 2100, 27–63.
- [8] Becker, S.C., Foster-Frey, J., Donovan, D.M., 2008. The phage K lytic enzyme LysK and lysostaphin act synergistically to kill MRSA. *FEMS Microbiol. Lett.* 287, 185–191.
- [9] Biller, J.A., King, S., Rosenthal, A., Grand, R.J., 1987. Efficacy of lactase-treated milk for lactose-intolerant pediatric patients. *J. Pediatr.* 111, 91–94.
- [10] Borysowski, J., Weber-Dabrowska, B., Gorski, A., 2006. Bacteriophage endolysins as a novel class of antibacterial agents. *Exp. Biol. Med.* (Maywood) 231, 366–377.
- [11] achumba, J.J., Antunes, F.A., Peres, G.F., Brumano, L.P., Santos, J.C., Da Silva, S.S., 2016. Current applications and different approaches for microbial l-asparaginase production. *Brazil. J. Microbiol.*: [Publication of the Brazilian Society for Microbiology] 47 (Suppl 1), 77–85.
- [12] Cantor, J.R., Panayiotou, V., Agnello, G., Georgiou, G., Stone, E.M., 2012. Engineering reduced-immunogenicity enzymes for amino acid depletion therapy in cancer. *Methods Enzymol.* 502, 291–319.

- [13] Cavuoto, P., Fenech, M.F., 2012. A review of methionine dependency and the role of methionine restriction in cancer growth control and life-span extension. *Cancer Treat. Rev.* 38, 726–736.
- [14] Chappi, D.M., Suresh, K.V., Patil, M.R., Desai, R., Tauro, D.P., Bharani, K.N.S.S., Parkar, M.I., Babaji, H.V., 2015. Comparison of clinical efficacy of methylprednisolone and serratiopeptidase for reduction of postoperative sequelae after lower third molar surgery. *J. Clin. Exp. Dent.* 7, e197–202.
- [15] Cheng, F., Yang, J., Schwaneberg, U., Zhu, L., 2019. Rational surface engineering of an arginine deiminase (an antitumor enzyme) for increased PEGylation efficiency. *Biotechnol. Bioeng.* 116, 2156–2166.
- [16] Cho, Y.H., Song, J.Y., Kim, K.M., Kim, M.K., Lee, I.Y., Kim, S.B., Kim, H.S., Han, N.S., Lee, B.H., Kim, B.S., 2010. Production of nattokinase by batch and fed-batch culture of *Bacillus subtilis*. *New Biotechnol.* 27, 341–346.
- [17] Dams, D., Briens, Y., 2019. Enzybiotics: Enzyme-Based Antibacterials as Therapeutics. *Adv. Exp. Med. Biol.* 1148, 233–253.
- [18] Daw, M.A., Falkiner, F.R., 1996. Bacteriocins: nature, function and structure. *Micron* 27, 467–479.
- [19] De Souza, P.M., De Oliveira Magalhaes, P., 2010. Application of microbial alpha-amylase in industry - A review. *Brazil. J. Microbiol.* [Publication of the Brazilian Society for Microbiology] 41, 850–861.
- [20] Dean, S.N., Turner, K.B., Medintz, I.L., Walper, S.A., 2017. Targeting and delivery of therapeutic enzymes. *Therapeut. Delivery* 8, 577–595.
- [21] Delage, B., Fennell, D.A., Nicholson, L., McNeish, I., Lemoine, N.R., Crook, T., Szlosarek, P.W., 2010. Arginine deprivation and argininosuccinate synthetase expression in the treatment of cancer. *Int. J. Cancer* 126, 2762–2772.
- [22] Dhankhar, R., Gupta, V., Kumar, S., Kapoor, R.K., Gulati, P., 2020. Microbial enzymes for deprivation of amino acid metabolism in malignant cells: biological strategy for cancer treatment. *Appl. Microbiol. Biotechnol.* 104, 2857–2869.
- [23] Diep, D.B., Nes, I.F., 2002. Ribosomally synthesized antibacterial peptides in Gram positive bacteria. *Curr. Drug Targets* 3, 107–122.
- [24] Diez-Martinez, R., De Paz, H.D., Garcia-Fernandez, E., Bustamante, N., Euler, C.W., Fischetti, V.A., Menendez, M., Garcia, P., 2015. A novel chimeric phage lysin with high in vitro and in vivo bactericidal activity against *Streptococcus pneumoniae*. *J. Antimicrobial Chemother.* 70, 1763–1773.
- [25] Droge, W., 2002. Free radicals in the physiological control of cell function. *Physiol. Rev.* 82, 47–95.
- [26] Egler, R.A., Ahuja, S.P., Matloub, Y., 2016. L-asparaginase in the treatment of patients with acute lymphoblastic leukemia. *J. Pharmacol. Pharmacotherapeut.* 7, 62–71.
- [27] Enzymes Market Size, Share & Trends Analysis Report By Application (Industrial Enzymes, Specialty Enzymes), By Product (Carbohydrase, Proteases, Lipases), By Source, By Region, And Segment Forecasts, 2020–2027. Report ID 978-1-68038- 022-4).
- [28] Ercan, D., Demirci, A., 2013. Production of human lysozyme in biofilm reactor and optimization of growth parameters of *Kluyveromyces lactis* K7. *Appl. Microbiol. Biotechnol.* 97, 6211–6221.
- [29] Ercan, D., Demirci, A., 2016. Recent advances for the production and recovery methods of lysozyme. *Crit. Rev. Biotechnol.* 36, 1078–1088.
- [30] Esch, P.M., Gerngross, H., Fabian, A., 1989. Reduction of postoperative swelling. Objective measurement of swelling of the upper ankle joint in treatment with serrapeptase— a prospective study. *Fortschr. Med.* 107 (67–68).
- [31] Fernandes, H.S., Silva Teixeira, C.S., Fernandes, P.A., Ramos, M.J., Cerqueira, N.M., 2017. Amino acid deprivation using enzymes as a targeted therapy for cancer and viral infections. *Expert Opin. Ther. Pat.* 27, 283–297.
- [32] Galvan Marquez, I.J., McKay, B., Wong, A., Cheetham, J.J., Bean, C., Golshani, A., Smith, M.L., 2020. Mode of action of nisin on *Escherichia coli*. *Can. J. Microbiol.* 66, 161–168.
- [33] Germolec, D.R., Shipkowski, K.A., Frawley, R.P., Evans, E., 2018. Markers of Inflammation. *Methods Mol. Biol.* 1803, 57–79.
- [34] Ghequire, M.G., Dillen, Y., Lambrechts, I., Proost, P., Wattiez, R., De Mot, R., 2015. Different Ancestries of R Tailocins in Rhizospheric *Pseudomonas* Isolates. *Genome Biol. Evolut.* 7, 2810–2828.
- [35] Glissen Brown, J.R., Singh, P., 2019. Coeliac disease. *Paediatr. Int. Child Health* 39, 23–31.
- [36] Griess, B., Tom, E., Domann, F., Teoh-Fitzgerald, M., 2017. Extracellular superoxide dismutase and its role in cancer. *Free Radical Biol. Med.* 112, 464–479.
- [37] Griffin, S.M., Alderson, D., Farndon, J.R., 1989. Acid resistant lipase as replacement therapy in chronic pancreatic exocrine insufficiency: a study in dogs. *Gut* 30, 1012–1015.
- [38] Gupte, V., Luthra, U., 2017. Analytical techniques for serratiopeptidase: A review. *J. Pharm. Anal.* 7, 203–207.
- [39] Han, R.Z., Xu, G.C., Dong, J.J., Ni, Y., 2016. Arginine deiminase: recent advances in discovery, crystal structure, and protein engineering for improved properties as an anti-tumor drug. *Appl. Microbiol. Biotechnol.* 100, 4747–4760.
- [40] Harding, J.J., Do, R.K., Dika, I.E., Hollywood, E., Uhlitskykh, K., Valentino, E., Wan, P., Hamilton, C., Feng, X., Johnston, A., Bomalaski, J., Li, C.F., O'Reilly, E.M., AbouAlfa, G.K., 2018. A phase 1 study of ADI-PEG 20 and modified FOLFOX6 in patients with advanced hepatocellular carcinoma and other gastrointestinal malignancies. *Cancer Chemother. Pharmacol.* 82, 429–440.
- [41] Hartig-Andreasen, C., Schroll, L., Lange, J., 2019. Clostridium histolyticum as first-line treatment of Dupuytren's disease. *Danish Med. J.* 66. M. Vachher et al. *Current Research in Biotechnology* 3 (2021) 195–208 205
- [42] Hoffman, R.M., Tan, Y., Li, S., Han, Q., Zavala Sr., J., Zavala Jr., J., 2019. Pilot Phase I Clinical Trial of Methioninase on High-Stage Cancer Patients: Rapid Depletion of Circulating Methionine. *Methods Mol. Biol.* 1866, 231–242.
- [43] Hoy, S.M., 2020. Collagenase Clostridium Histolyticum: A Review in Peyronie's Disease. *Clin. Drug Invest.* 40, 83–92.
- [44] Ianiro, G., Bibbo, S., Gasbarrini, A., Cammarota, G., 2014. Therapeutic modulation of gut microbiota: current clinical applications and future perspectives. *Curr. Drug Targets* 15, 762–770.
- [45] Ianiro, G., Pecere, S., Giorgio, V., Gasbarrini, A., Cammarota, G., 2016. Digestive Enzyme Supplementation in Gastrointestinal Diseases. *Curr. Drug Metab.* 17, 187–193.
- [46] Ishihara, T., Tanaka, K., Tasaka, Y., Namba, T., Suzuki, J., Okamoto, S., Hibi, T., Takenaga, M., Igarashi, R., Sato, K., Mizushima, Y., Mizushima, T., 2009. Therapeutic effect of lecithinized superoxide dismutase against colitis. *J. Pharmacol. Exp. Therapeut.* 328, 152–164.
- [47] Jadhav, S.B., Shah, N., Rathi, A., Rathi, V., 2020. Serratiopeptidase: Insights into the therapeutic applications. *Biotechnol. Rep. (Amst)* 28, e00544.
- [48] Jayasekara, S., Ratnayake, R., 2019. Microbial cellulases: an overview and applications. *Cellulose* 22.
- [49] Jesuraj, S.A.V., Sarker, M.M.R., Ming, L.C., Praya, S.M.J., Ravikumar, M., Wui, W.T., 2017. Enhancement of the production of L-glutaminase, an anticancer enzyme, from *Aeromonas veronii* by adaptive and induced mutation techniques. *PLoS ONE* 12, e0181745.
- [50] Juturu, V., Wu, J.C., 2018. Microbial production of bacteriocins: Latest research development and applications. *Biotechnol. Adv.* 36, 2187–2200.
- [51] Kasinathan, N., Volety, S.M., Josyula, V.R., 2016. Chondroitinase: A promising therapeutic enzyme. *Crit. Rev. Microbiol.* 42, 474–484.
- [52] Kaur, J., Singh, P., Sharma, D., Harjai, K., Chhibber, S., 2020. A potent enzymatic against methicillin-resistant *Staphylococcus aureus*. *Virus Genes* 56, 480–497.
- [53] Klasen, H.J., 2000. A review on the nonoperative removal of necrotic tissue from burn wounds. *Burns: J. Int. Soc. Burn Injuries* 26, 207–222.
- [54] Knott, S.R.V., Wagenblast, E., Khan, S., Kim, S.Y., Soto, M., Wagner, M., Turgeon, M.O., Fish, L., Erard, N., Gable, A.L., Maceli, A.R., Dickopf, S., Papachristou, E.K., D'Santos, C.S., Carey, L.A., Wilkinson, J.E., Harrell, J.C., Perou, C.M., Goodarzi, H., Poulgiannis, G., Hannon, G.J., 2018. Asparagine bioavailability governs metastasis in a model of breast cancer. *Nature* 554, 378–381.
- [55] Li, M., 2018. Enzyme Replacement Therapy: A Review and Its Role in Treating Lysosomal Storage Diseases. *Pediatr. Ann.* 47, e191–e197.

- [56] Sheets, A.R., Demidova-Rice, T.N., Shi, L., Ronfard, V., Grover, K.V., Herman, I.M., 2016. Identification and Characterization of Novel Matrix-Derived Bioactive Peptides: A Role for Collagenase from Santyl(R) Ointment in Post-Debridement Wound Healing?. PLoS ONE 11, e0159598.
- [57] Srivastava, V., Mishra, S., Chaudhuri, T.K., 2019. Enhanced production of recombinant serratiopeptidase in *Escherichia coli* and its characterization as a potential biosimilar to native biotherapeutic counterpart. Microb. Cell Fact. 18, 215.
- [58] Steiger, S., Harper, J.L., 2014. Mechanisms of spontaneous resolution of acute gouty inflammation. Curr. Rheumatol. Rep. 16, 392.
- [59] Stevens, J., Wyatt, C., Brown, P., Patel, D., Grujic, D., Freedman, S.D., 2018. Absorption and Safety With Sustained Use of RELIZORB Evaluation (ASSURE) Study in Patients With Cystic Fibrosis Receiving Enteral Feeding. J. Pediatr. Gastroenterol. Nutr. 67, 527–532.
- [60] Tabe, Y., Lorenzi, P.L., Konopleva, M., 2019. Amino acid metabolism in hematologic malignancies and the era of targeted therapy. Blood 134, 1014–1023.
- [61] Taipa, M.A., Fernandes, P., de Carvalho, C., 2019. Production and Purification of Therapeutic Enzymes. Adv. Exp. Med. Biol. 1148, 1–24.
- [62] Tamimi, Z., Al-Habashneh, R., Hamad, I., Al-Ghazawi, M.A., Roqaa, A.A., Kharashgeh, H., 2021. Efficacy of serratiopeptidase after impacted third molar surgery: a randomized controlled clinical trial. BMC Oral Health 21, 91.
- [63] Taneja, K., Bajaj, B.K., Kumar, S., Dilbaghi, N., 2017. Production, purification and characterization of fibrinolytic enzyme from *Serratia* sp. KG-2-1 using optimized media. 3. Biotech 7, 184.
- [64] Tiwari, M., 2017. The role of serratiopeptidase in the resolution of inflammation. Asian J. Pharm. Sci. 12, 209–215.
- [65] UmaMaheswari, T., Hemalatha, T., Sankaranarayanan, P., Puvanakrishnan, R., 2016. Enzyme Therapy: Current Perspectives. Indian J. Exp. Biol. 54, 7–16.
- [66] Vachher, M., Arora, K., Burman, A., Kumar, B., 2020. NAMPT, GRN, and SERPINE1 signature as predictor of disease progression and survival in gliomas. J. Cell. Biochem. 121, 3010–3023.
- [67] Vakili, B., Nezafat, N., Negahdaripour, M., Yari, M., Zare, B., Ghasemi, Y., 2017. Staphylokinase Enzyme: An Overview of Structure, Function and Engineered Forms. Curr. Pharm. Biotechnol. 18, 1026–1037.
- [68] Vijayaraghavan, P., Arasu, M.V., AnanthaRajan, R., Al-Dhabi, N.A., 2019. Enhanced production of fibrinolytic enzyme by a new *Xanthomonas oryzae* IND3 using lowcost culture medium by response surface methodology. Saudi J. Biol. Sci. 26, 217–224.
- [69] Weng, Y., Yao, J., Sparks, S., Wang, K.Y., 2017. Nattokinase: An Oral Antithrombotic Agent for the Prevention of Cardiovascular Disease. Int. J. Mol. Sci. 18.
- [70] Xiong, L., Teng, J.L., Botelho, M.G., Lo, R.C., Lau, S.K., Woo, P.C., 2016. Arginine Metabolism in Bacterial Pathogenesis and Cancer Therapy. Int. J. Mol. Sci. 17, 363.
- [71] M. Vachher et al. Current Research in Biotechnology 3 (2021) 195–208 207
- [72] Xu, L., Liang, G., Chen, B., Tan, X., Xiang, H., Liao, C., 2020. A Computational Method for the Identification of Endolysins and Autolysins. Protein Pept. Lett. 27, 329–336.
- [73] Yang, J., Tao, R., Wang, L., Song, L., Wang, Y., Gong, C., Yao, S., Wu, Q., 2019. Thermosensitive Micelles Encapsulating Phenylalanine Ammonia Lyase Act as a Sustained and Efficacious Therapy Against Colorectal Cancer. J. Biomed. Nanotechnol. 15, 717–727.
- [74] Yari, M., Ghoshoon, M.B., Vakili, B., Ghasemi, Y., 2017. Therapeutic Enzymes: Applications and Approaches to Pharmacological Improvement. Curr. Pharm. Biotechnol. 18, 531–540.
- [75] Zarei, M., Rahbar, M.R., Morowvat, M.H., Nezafat, N., Negahdaripour, M., Berenjian, A., Ghasemi, Y., 2019. Arginine Deiminase: Current Understanding and Applications. Recent Pat. Biotechnol. 13, 124–136.

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